



# ENM TUTORIALS

How to use the Pathway module of ArrayAnalysis.org for pathway analysis of microarray data

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VERSION:	V.1.0.
MAIN AUTHOR:	Friederike Ehrhart
PARTNER:	UM
CONTACT DETAILS:	<a href="mailto:friederike.ehrhart@maastrichtuniversity.nl">friederike.ehrhart@maastrichtuniversity.nl</a> <a href="mailto:linda.rieswijk@maastrichtuniversity.nl">linda.rieswijk@maastrichtuniversity.nl</a> <a href="mailto:egon.willighagen@maastrichtuniversity.nl">egon.willighagen@maastrichtuniversity.nl</a> +31(0)43-38 82913
AUTHORS:	Lars Eijssen, Anwasha Bohler, Linda Rieswijk, Egon Willighagen, Penny Nymark
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# TABLE OF CONTENTS

## [1. INTRODUCTION](#)

## [2. APPLICATION DETAILS](#)

[First step: load the data file and select species](#)

[Second step: Identifier Mapping](#)

[Third step: Set criterion for z-score calculation](#)

[Fourth step: Creating a visualization](#)

[Execution](#)

[Results](#)

[Interpret the results of the Path module](#)

## [3. ACKNOWLEDGMENTS](#)

## [4. REFERENCES](#)

## [5. KEYWORDS](#)

# 1. INTRODUCTION

PathVisio and ArrayAnalysis.org are open source, free to use online platforms for analysis of microarray data - and an alternative program for Chipster. This tutorial shows how to use the Path module (Pathway module, PathVisio webtool) of ArrayAnalysis which is designed for doing pathway analysis on microarray data. All source code has been written in R and is available at [https://github.com/BiGCAT-UM/Path\\_Module](https://github.com/BiGCAT-UM/Path_Module).

This technical documentation has two main objectives:

- to guide you in the use of the Path module
- to give interpretative help on the outputs of the module

The Path module can be run :

- on-line via the <http://www.arrayanalysis.org> webportal (follow "Get started" and choose "Pathway analysis")
- or as an automated R workflow from a local computer

The main functions of the Path module are:

- to import a dataset;
- to create a visualization;
- to calculate z-scores based on the criterion;
- to return a list of pathways sorted on the basis of z-scores.

How to use the documentation: As shown in the Table Of Contents, you will find the separate sections :

- Using the on-line Path module
- Interpreting the results provided.

Bug tracking system: If you encounter an issue by using the code, you can report it at any moment on our internal tracking system : <http://trac.bigcat.unimaas.nl/arrayanalysis/newticket>. You can also use this system to post comments or feature suggestions.

Example gene level statistics file: An example dataset is available. When running the module, you can check a box to use this data set (Example1) in order to explore the functionality of the module.

# 2. APPLICATION DETAILS

You can access the on-line module on [arrayanalysis.org](http://arrayanalysis.org) webportal: (follow "Get started" and choose "Statistical analysis"). You don't need to log in; you just need to prepare a gene level statistics file containing the statistical contrasts between the different groups of your Affymetrix .CEL files (you may also obtain the file by running the statistical analysis module).

The on-line module contains four steps before the launch of the analysis:

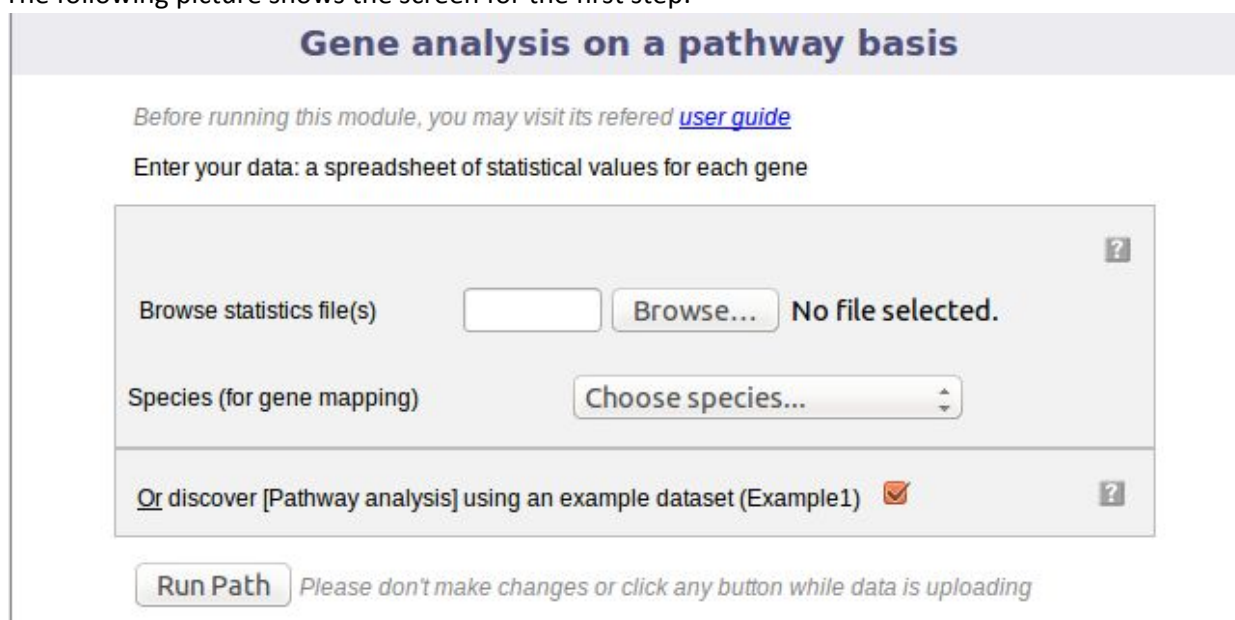
- [Step1](#): First you load the gene level statistics file and select species. Alternatively, you might select the Example dataset for exploring the module. In that case you do not need to select the species. The dataset used is for Human. Click on Run Path, to proceed.
- [Step2](#): Choose the column in the data file containing the identifiers and the database used for annotation if all the identifiers are from the same database. If different identifier systems are used for annotating the dataset, the system code column has to be chosen. The system code specifies which database each identifier belongs to.
- [Step3](#): Specify a criterion for calculating the z-scores.
- [Step4](#): Select color criteria for visualization of the uploaded data on the pathways.

Then:

- [Execution](#): The module is executed with the settings you choose
- [Results](#): You get the results after the execution step, at the website or by e-mail.

### FIRST STEP: LOAD THE DATA FILE AND SELECT SPECIES

The following picture shows the screen for the first step:



**Gene analysis on a pathway basis**

Before running this module, you may visit its referred [user guide](#)

Enter your data: a spreadsheet of statistical values for each gene

Browse statistics file(s)   No file selected.

Species (for gene mapping)

Or discover [Pathway analysis] using an example dataset (Example1)

Please don't make changes or click any button while data is uploading

This dialog allows you to upload a tab-delimited text file with (gene level statistics) data and choose the relevant species. Alternatively, the module can be run with an example data set, by ticking the checkbox presented. The interrogation mark button will give you contextual help.

### SECOND STEP: IDENTIFIER MAPPING

The following part of the online form is used for the second step:

### Identifier Mapping

Define Annotation Columns ?

Choose the column containing identifiers and choose a database or a system code column for mapping identifiers.

Identifier Column	<input type="text" value="ENSG_ID"/>
Database	<input type="text" value="Choose database..."/>
System Code Column	<input type="text"/>

Your dataset has been uploaded. For mapping the uploaded data to the pathways, the annotation information needs to be filled in.

"Identifier Column" Choose the column in the uploaded data file containing the identifiers used for annotation.

"Database" If the identifiers used for annotation are all from the same database, then select the database.

OR "System Code" If identifiers from different databases are used for annotation then the a column containing the [system code](#) of the database needs to be selected.

The interrogation mark button will give you contextual help.

### THIRD STEP: SET CRITERION FOR Z-SCORE CALCULATION

The following part of the online form is used for the third step:

**Pathway statistics** Define a criterion for selection of relevant pathways. ?

A statistical test under the hypergeometric distribution (**Z score**) is calculated for each pathway; a high Z score indicates that the pathway has more genes that pass the criterion than expected by chance.

Expression:  Reset

Variables	Operators
ENSG_ID SysCode logFC FoldChange AveExpr t P.Value adj.P.Val B	AND OR = < > <= >= <>

Select a criterion for calculating the z-score. You could, e.g. specify a criterion based on a fold change threshold. You can either type the expression in the "Expression" field or you can use the available parameters and operators listed by clicking on them.

#### FOURTH STEP: CREATING A VISUALIZATION

The following part of the online form is used for the fourth step:

### Color rules ?

A rule is defined by one color corresponding to one expression, *for example* If the P.value is below 0.01, then color green.

color set 1 Delete Selected Rule Add Rule

00FF00 [P.Value] < 0.05 Reset

Delete Selection
Add Color Set

#### Variables

ENSG\_ID  
 SysCode  
 logFC  
 FoldChange  
 AveExpr  
 t  
 P.Value  
 adj.P.Val  
 B

#### Operators

AND  
 OR  
 =  
 <  
 >  
 <=  
 >=  
 <>

### Gradient color set ?

A color gradient is defined by colors and corresponding values. In the example gradient, proposed as default settings, a color gradient is applied to logFC. In the gradient blue corresponds to logFC= -1, white corresponds to logFC= 0 and red corresponds to logFC= +1.

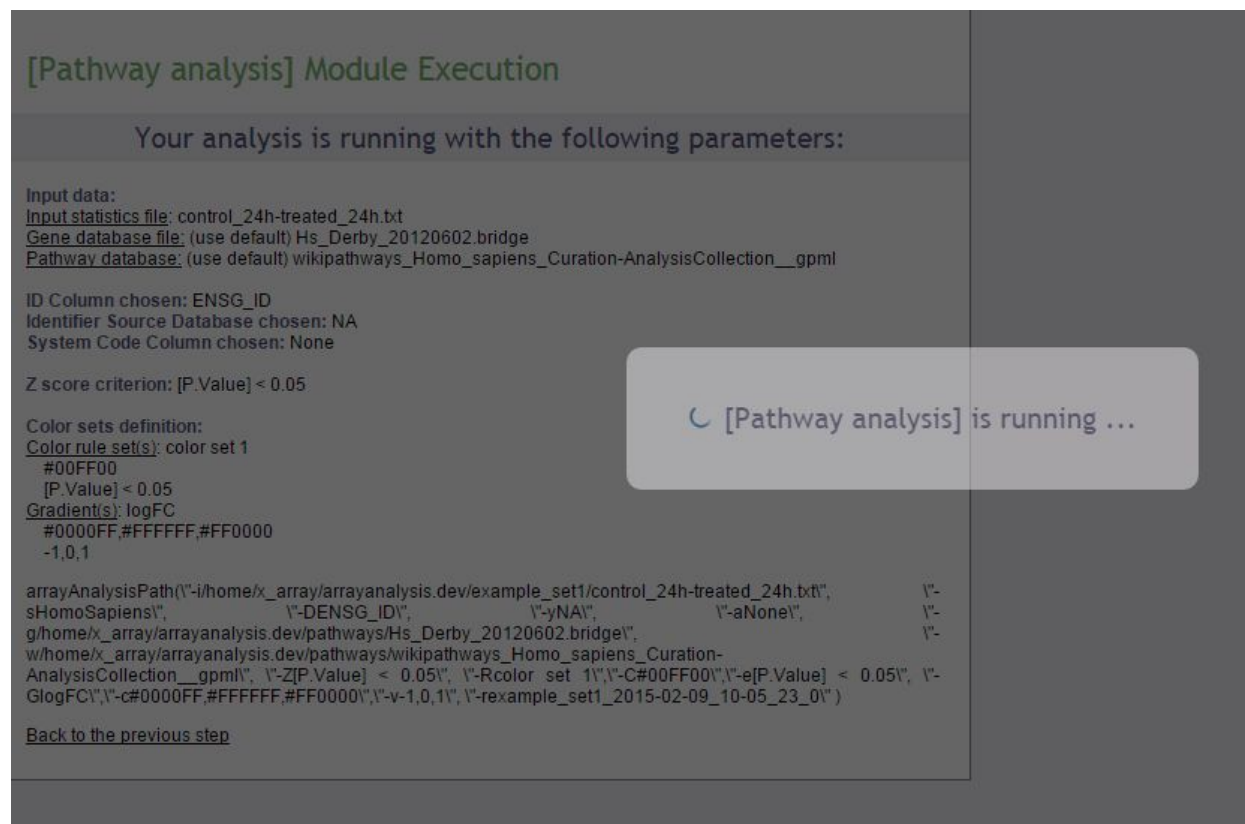
Column Name	Color<-->Value	Color<-->Value	Color<-->Value
<input type="checkbox"/> logFC	<span style="background-color: #0000FF; color: white; padding: 2px;">0000FF</span> -1	<span style="background-color: #FFFFFF; color: black; padding: 2px;">FFFFFF</span> 0	<span style="background-color: #FF0000; color: white; padding: 2px;">FF0000</span> 1

Delete Selection
Add Gradient

Data can be visualized on pathways using colours. A gradient colouring scheme can be used to visualize a range of data on a gene (e.g. fold change) while a rule can be applied for certain criteria allowing only the genes which qualify to be coloured (e.g. P Value "<" 0.05)

## EXECUTION

After clicking 'Run' the module is executed.



**[Pathway analysis] Module Execution**

Your analysis is running with the following parameters:

**Input data:**  
Input statistics file: control\_24h-treated\_24h.txt  
Gene database file: (use default) Hs\_Derby\_20120602.bridge  
Pathway database: (use default) wikipathways\_Homo\_sapiens\_Curation-AnalysisCollection\_\_gpml

ID Column chosen: ENSG\_ID  
Identifier Source Database chosen: NA  
System Code Column chosen: None

Z score criterion: [P.Value] < 0.05

**Color sets definition:**  
Color rule set(s): color set 1  
#00FF00  
[P.Value] < 0.05  
Gradient(s): logFC  
#0000FF,#FFFFFF,#FF0000  
-1,0,1

arrayAnalysisPath("i:/home/x\_array/arrayanalysis.dev/example\_set1/control\_24h-treated\_24h.txt", "sHomoSapiens", "D-ENSG\_ID", "y-NA", "a-None", "g/home/x\_array/arrayanalysis.dev/pathways/Hs\_Derby\_20120602.bridge", "w/home/x\_array/arrayanalysis.dev/pathways/wikipathways\_Homo\_sapiens\_Curation-AnalysisCollection\_\_gpml", "Z[P.Value] < 0.05", "Rcolor set 1", "C-#00FF00", "e[P.Value] < 0.05", "GlogFC", "c-#0000FF,#FFFFFF,#FF0000", "v-1,0,1", "r-example\_set1\_2015-02-09\_10-05\_23\_0")

[Back to the previous step](#)

[Pathway analysis] is running ...

## RESULTS

Upon completion a page of results is displayed on your screen.



## [Pathway analysis] Module Execution

Your analysis is running with the following parameters:

### Input data:

Input statistics file: control\_24h-treated\_24h.txt

Gene database file: (use default) Hs\_Derby\_20120602.bridge

Pathway database: (use default) wikipathways\_Homo\_sapiens\_Curation-AnalysisCollection\_\_gpml

ID Column chosen: ENSG\_ID

Identifier Source Database chosen: En

System Code Column chosen: None

Z score criterion: [P.Value] < 0.05

### Color sets definition:

Color rule set(s): color set 1

#00FF00

[P.Value] < 0.05

Gradient(s): logFC

#0000FF,#FFFFFF,#FF0000

-1,0,1

```
arrayAnalysisPath("-i/home/x_array/arrayanalysis.dev/example_set1/control_24h-treated_24h.txt", \n                 sHomoSapiens", \n                 "-DENSG_ID", \n                 "-yEn", \n                 "-aNone", \n                 \n                 g/home/x_array/arrayanalysis.dev/pathways/Hs_Derby_20120602.bridge", \n                 \n                 w/home/x_array/arrayanalysis.dev/pathways/wikipathways_Homo_sapiens_Curation- \n                 AnalysisCollection__gpml", \n                 "-Z[P.Value] < 0.05", \n                 "-Rcolor set 1", \n                 "-C#00FF00", \n                 "-e[P.Value] < 0.05", \n                 "-GlogFC", \n                 "-c#0000FF,#FFFFFF,#FF0000", \n                 "-v-1,0,1", \n                 "-rexample_set1_2015-02-09_10-10_44_0")
```

[Back to the previous step](#)

```
cd /home/x_array/arrayanalysis.dev//Path_Module/
Rscript /home/x_array/arrayanalysis.dev/temp/launchFilePath_example_set1_2015-02-09_10-10_44_0.R

cd /home/x_array/arrayanalysis.dev/temp/example_set1_2015-02-09_10-10_44_0_Path/
zip -r pathways_results_example_set1_2015-02-09_10-10_44_0.zip contents index.html
```

## Results for example\_set1\_2015-02-09\_10-10\_44\_0:

Result files (Right click on the following link(s) to save the corresponding file)

[Open log file](#) containing standard output, warning and error messages from the execution.  
You may also consult this text file on the following section: Output message (STDOUT & STDERR).

[Download zip file](#) with result html pages.

[Result page](#) Open the complete result page in a new tab or window

Summary tables for relevant pathways:

Powered by [PathVisio](#)

Gene/Protein found in pathways (n): 3252

Gene/Protein meeting criterion (r): 892

Twenty more significant pathways	n	r	z
<a href="#">Apoptosis Modulation and Signaling</a>	76	35	3.68
<a href="#">Serotonin Receptor 2 -&gt; ELK-SRF/GATA4 signaling</a>	15	9	2.83
<a href="#">Cholesterol Biosynthesis</a>	13	8	2.76
<a href="#">Serotonin Receptor 4/6/7 -&gt; NR3C signaling</a>	17	9	2.36
<a href="#">mRNA processing</a>	116	42	2.16
<a href="#">Proteasome Degradation</a>	58	23	2.11
<a href="#">FAS pathway and Stress induction of HSP regulation</a>	35	15	2.06
<a href="#">Eukaryotic Transcription Initiation</a>	35	15	2.06
<a href="#">MAPK signaling pathway</a>	152	52	1.92
<a href="#">Fatty Acid Biosynthesis</a>	22	10	1.90
<a href="#">IL-6 Signaling Pathway</a>	92	33	1.84
<a href="#">Translation Factors</a>	40	16	1.79
<a href="#">Apoptosis</a>	80	29	1.79

<a href="#">IL-6 Signaling Pathway</a>	92	33	1.84
<a href="#">Translation Factors</a>	40	16	1.79
<a href="#">Apoptosis</a>	80	29	1.79
<a href="#">Pentose Phosphate Pathway</a>	7	4	1.76
<a href="#">Acetylcholine Synthesis</a>	7	4	1.76
<a href="#">Keap1-Nrf2</a>	12	6	1.76
<a href="#">Cell cycle</a>	84	30	1.72
<a href="#">Oxidative Stress</a>	26	11	1.71
<a href="#">Folic Acid Network</a>	26	11	1.71
<a href="#">Fluoropyrimidine Activity</a>	29	12	1.69
<a href="#">more pathways...</a>			

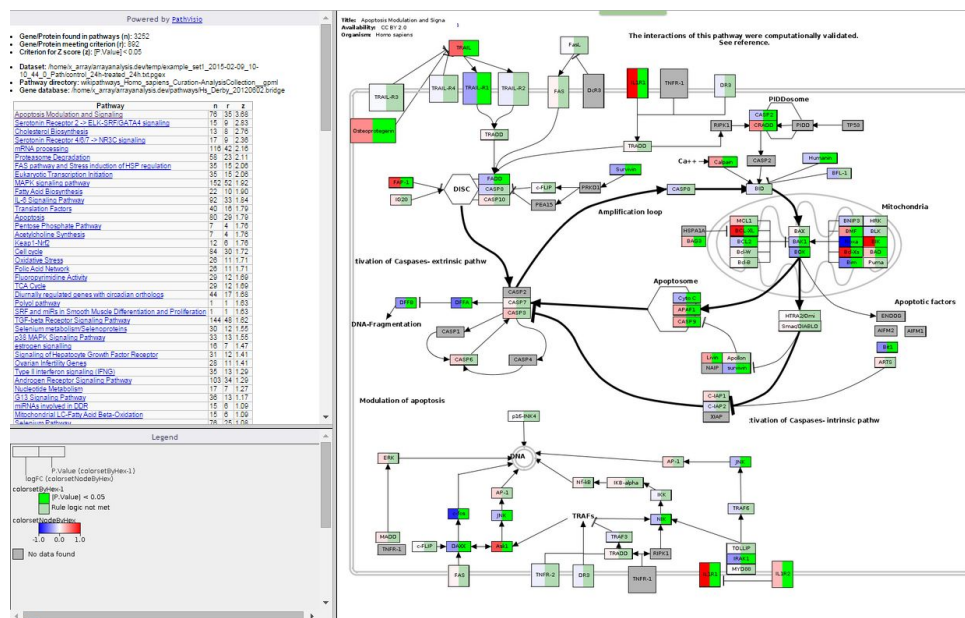
### Output message (STDOUT & STDERR):

#### Standard output:

```
[1] "Parameters have been registered"
[1] "Libraries are loaded"
[1] "Importing Data ...."
[1]      "/home/x_array/arrayanalysis.dev/temp/example_set1_2015-02-09_10-10_44_0_Path/control_24h-
treated_24h.txt.pgexfile created!"
[1] "Creating visualisation..."
[1]      "/home/x_array/arrayanalysis.dev/temp/example_set1_2015-02-09_10-10_44_0_Path/control_24h-
treated_24h.txt.pgex.xml-visualization file created!"
[1] "Calculating Pathway Statistics and Exporting results..."
[1]      "Pathway                Collection                used:
/home/x_array/arrayanalysis.dev/pathways/wikipathways_Homo_sapiens_Curation-AnalysisCollection__gpml/"
[1]      "Gene                Identifier                Mapping                Database                Used:
/home/x_array/arrayanalysis.dev/pathways/Hs_Derby_20120602.bridge"
[1]      "Dataset Imported:      /home/x_array/arrayanalysis.dev/temp/example_set1_2015-02-09_10-
10_44_0_Path/control_24h-treated_24h.txt.pgex"
[1] "Z score criterion: [P.Value] < 0.05"
[1] "Result Directory on Calculation server /home/x_array/arrayanalysis.dev/temp/example_set1_2015-02-
09_10-10_44_0_Path"
[1] "Run completed"
```

In the first part of the screen, your settings are recalled. Then links to the log file of the run and to the zip file containing all results (index file, pathway images, and related backpages) are presented. The results will be described in the next section of this documentation.

## INTERPRET THE RESULTS OF THE PATH MODULE



The output consists of :

An index file in html format, which contains:

- The number of genes/proteins in the dat aset found in the pathways of the pathway collection.
- The number of genes/proteins out of the above that meet the user defined criterion for z-score calculation.
- The criterio used for z-score calculation.
- The dataset used.
- The pathway directory used.
- The identifier mapping database used.
- A clickable list of all the pathways on which the genes of interest have been visualized sorted on the basis of z-scores.
- The file also contains the legend to better understand the visualization.

A contents folder: contains the backpages for all the pathways containing gene expression information, the legend file, the statistics file and all the colour coded pathway images.

WikiPathways provides a portal for nanomaterial relevant pathway information:

<http://www.wikipathways.org/index.php/Portal:Nanomaterials>

## 3. ACKNOWLEDGMENTS

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## 4. REFERENCES

We would like to express our gratitude for using the open-access applications of ArrayAnalysis.org. This tutorial is derived from <http://www.arrayanalysis.org/> documentation originally written by Lars Eijssen and Anwasha Bohler.

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Adding automated Statistical Analysis and Biological Evaluation modules to [www.arrayanalysis.org](http://www.arrayanalysis.org). Master's thesis of Anwasha Bohler (Dutta) Advisor : Lars M.T. Eijssen, doi: 10.13140/2.1.5150.6244 [PathVisio 3: An Extendable Pathway Analysis Toolbox](#), Kutmon M, van Iersel MP, Bohler A, Kelder T, Nunes N, Pico AR, and Evelo, CT, PLoS Computational Biology (2015) 11(2): e1004085, doi: [10.1371/journal.pcbi.1004085](https://doi.org/10.1371/journal.pcbi.1004085), pubmed: [25706687](https://pubmed.ncbi.nlm.nih.gov/25706687/)

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## 5. KEYWORDS

Microarray data analysis

Statistics

Systems biology

Pathway and network analysis